

Comparing Form Bio Data Analysis Performance With and Without NVIDIA Clara Parabricks



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Abstract

Genetic variant detection has wide application in genomics including identifying genetic changes that drive phenotypic changes. The detection of differences between a genomic re-sequencing experiment allows researchers to detect genetic diversity with a population, identify disease causing genetic changes and discover genetic markers of speciation when comparing closely related species. Germline variant detection is used to detect inherited genetic changes, whereas somatic variant detection is used to detect acquired genetic changes including the genetic mutations that drive tumor growth. There are many types of genetic variation including single nucleotide variants (SNVs) and insertions/deletions (indels), copy number variants (CNVs), and structural variants (SVs), which include large insertions, deletions, inversions, duplications and translocations. Over the last decade several open source tools have been developed to improve the accuracy and

speed of SNV/Indel variant detection including BWA mem, minimap2, GATK and DeepVariant. However it can still take days or weeks with these tools to analyze whole genome sequencing without very large computational clusters so that analysis runs in parallel over many machines. Accelerated versions of these tools using GPUs has been shown to reduce computational time and resources. We analyzed a publicly available dataset using DNA from a sample named Genome in a Bottle (GIAB), NA12878 to determine the sensitivity and specificity of running NVIDIA Parabricks compared to the open source versions of the same tools on Google Cloud Platform. The resulting analysis using NVIDIA Parabricks was faster and cheaper than using open source tools. The optimization did not significantly change the underlying alignments produced. Sensitivity and specificity were similar to the open source version of the tools even when only the alignment was used.

Methods

The workflow “Genomics: Germline Variant Analysis” determines genetic variants including SNVs, insertions and deletions of high-quality NGS data compared to a reference genome. Reads are trimmed using TrimGalore¹⁻³, to trim low quality (qual < 25) ends of reads and remove reads < 35bp. This workflow can be run with native open-source tools (NOST) or with Parabricks.

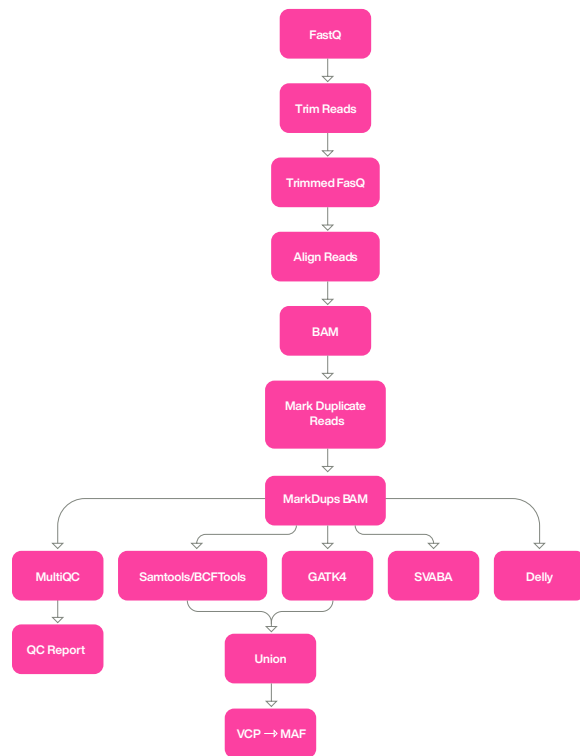
With NOST, trimmed reads are aligned to a reference genome using BWA mem⁴ or Minimap²⁵. Duplicates reads can optionally be marked using Picard MarkDuplicates⁶. BAMs from the same sample

generated by multiple runs are merged using samtools⁷. Alignment quality is assessed using FastQC2, Samtools⁷, Bedtools⁸. With Parabricks, trimmed reads are aligned, duplicate reads are marked and alignment quality is accessed using fq2bam^{9,10}. Quality metrics are summarized with MultiQC. With NOST, variants can be detected with joint calling using Freebayes¹¹, Samtools/Bcftools⁷ and GATK¹². With Parabricks, variants can be detected with GATK¹² and DeepVariant¹³ to produce gVCF files. Genotyping of gVCF files is determined using¹⁴. Variants effects are determined using SNPEff¹⁵.

Workflow Diagrams



With NVIDIA Parabricks



Native open-source tools

Results

We ran our workflow using open source versions of alignment and variant calling and with Parabricks with 3 separate sequencing runs of NA12878 as separate samples spanning 20M-160M reads. The full workflow ran in 344 minutes using Parabricks and 2,793 minutes using open source tools with multiple steps running it parallel, representing an 88% time savings. The biggest increase in efficiency came from variant calling where the median job took 1,376 minutes for open source tools and 94 minutes using Parabricks, representing a 93% time savings. The cost of running with Parabricks was US\$45 vs US\$93 with open source tools, representing a 52% savings. The resulting variants have comparable sensitivity, within 3%, for variant calling methods that use Parabricks including GATK and DeepVariant and those that do not have an accelerated version such as Freebayes, Samtools.

| SAMPLE ID | METHOD | SN | PPV |
|-----------|-------------|------|------|
| NOST | gatk | 95.2 | 94.8 |
| NP | gatk | 92.2 | 93.8 |
| NOST | deepvariant | 91.0 | 98.9 |
| NP | deepvariant | 87.8 | 99.6 |

References

- Martin, M. Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet journal* 17, 10 (2011).
- Andrews, S. et al. FastQC. (2012).
- Krueger, F., James, F., Ewels, P., Afyounian, E. & Schuster-Boeckler, B. FelixKrueger/TrimGalore: v0.6.7 - DOI via Zenodo. (2021) doi:10.5281/ZENODO.5127899.
- Li, H. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *ArXiv Prepr. ArXiv 00*, 3 (2013).
- Li, H. Minimap2: pairwise alignment for nucleotide sequences. *Bioinformatics* 34, 3094–3100 (2018).
- Thomer, A. K., Twidale, M. B., Guo, J. & Yoder, M. J. Picard Tools. in *Conference on Human Factors in Computing Systems - Proceedings* (2016).
- Li, H. et al. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25, 2078–2079 (2009).
- Quinlan, A. R. & Hall, I. M. BEDTools: A flexible suite of utilities for comparing genomic features. *Bioinformatics* 26, 841–842 (2010).
- Franke, K. R. & Crowgey, E. L. Accelerating next generation sequencing data analysis: an evaluation of optimized best practices for Genome Analysis Toolkit algorithms. *Genomics Inform.* 18, e10 (2020).
- Friedman, S., Gauthier, L., Farjoun, Y. & Banks, E. Lean and deep models for more accurate filtering of SNP and INDEL variant calls. *Bioinformatics* 36, 2060–2067 (2020).
- Garrison, E. & Marth, G. Haplotype-based variant detection from short-read sequencing. (2012).
- DePristo, M. A. et al. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nat. Genet.* 43, 491–498 (2011).
- Poplin, R. et al. A universal SNP and small-indel variant caller using deep neural networks. *Nat. Biotechnol.* 36, 983–987 (2018).
- Lin, M. F. et al. GLnexus: joint variant calling for large cohort sequencing. <http://biorxiv.org/lookup/doi/10.1101/343970> (2018) doi:10.1101/343970.
- Cingolani, P. et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff. *Fly (Austin)* 6, 80–92 (2012).